For Research Use

TakaRa

SYBR® *Premix Ex Taq*™ (Tli RNaseH Plus)

Product Manual





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I. Description

SYBR *Premix Ex Taq* (Tli RNaseH Plus) is designed for intercalator-based real-time PCR using SYBR Green I*. It is supplied at a 2X concentration and includes SYBR Green I at a concentration appropriate for real time monitoring, making it easy to prepare reaction mixtures. The 2X premixed reagent also contains Tli RNase H, a heat-resistant RNase H that minimizes PCR inhibition by degrading residual mRNA when using cDNA as template.

The combination of *TaKaRa Ex Taq*® HS, a hot start PCR enzyme that includes an anti-*Taq* antibody, and a buffer optimized for real-time PCR, allows high amplification efficiency and detection sensitivity. This product is suitable for high-speed PCR and enables accurate assay and detection of targets, making it possible to obtain highly reproducible and reliable real-time PCR results.

Benefits:

- (1) Allows rapid and accurate detection and assay of targets by real-time PCR.
- (2) 2X concentration premixed with SYBR Green I; simply add primers, template, and sterile distilled water to perform intercalator-based real-time PCR.
- (3) TaKaRa Ex Taq HS, a hot start PCR enzyme, is used for PCR. The buffer system has been optimized for real-time PCR, allowing good amplification efficiency and high-sensitivity detection.
- (4) The 2X reagent is premixed with Tli RNase H, a heat-resistant RNase H that minimizes PCR inhibition by residual mRNA when using cDNA as template.
- *: Takara Bio is under a license agreement with Molecular Probes Inc. for the use of SYBR Green I as a reagent for research purposes.

II. Principle

This product uses *TaKaRa Ex Taq* HS for PCR amplification. PCR amplification products may be monitored in real time using SYBR Green I.

1. PCR

URL:http://www.takara-bio.com

PCR is a technique used to amplify specific target sequences from minute amounts of DNA. By repeating three cycles of heat denaturation, primer annealing, and primer extension, the target fragment is amplified up to a million times by DNA polymerase within a short time.

This product uses *TaKaRa Ex Taq* HS, a hot start PCR enzyme that prevents non-specific amplification resulting from mispriming or primer dimer formation during reaction mixture preparation or other pre-cycling steps thereby allowing high-sensitivity detection.

2. Fluorescent detection - Intercalator method

This method uses a DNA intercalator (e.g., SYBR Green I) that emits fluorescence when bound to double-strand DNA. Monitoring fluorescence allows for quantification of amplification products.

Measuring the fluorescence intensity also provides the melting temperature of amplified DNA.

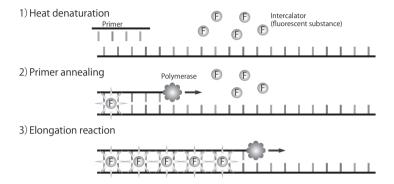


Figure 1. Fluorescent intercalator detection method.

III. Kit Components [for 40 x 50 μ l reactions]

SYBR <i>Premix Ex Taq</i> (Tli RNaseH Plus) (2X conc.)*1	1 ml
ROX Reference Dye (50X conc.)*2	40 µl
ROX Reference Dye II (50X conc.)*2	40 µ1

- *1: Contains *TaKaRa Ex Taq* HS, dNTP Mixture, Mg²⁺, Tli RNase H, and SYBR Green I. (For this product's quality specifications, please refer to Section X.)
- *2: ROX Reference Dye is used for analyses with devices that correct for betweenwell fluorescent signal, such as the real-time PCR devices by Life Technologies.
 - ◆ Use ROX Reference Dye
 - Applied Biosystems 7300 Real-Time PCR System
 - Applied Biosystems StepOnePlus Real-Time PCR System (Life Technologies)
 - ◆ Use ROX Reference Dye II
 - Applied Biosystems 7500/7500 Fast Real-Time PCR System (Life Technologies)
 - ◆ Do not use this component
 - Thermal Cycler Dice™ Real Time System Single (Cat. #TP850/TP870)
 Thermal Cycler Dice Real Time System Lite (Cat. #TP700/TP760)
 - Smart Cycler II System*3
 - LightCycler/LightCycler 480 System (Roche Diagnostics)
 - CFX96 Real-Time PCR Detection System (Bio-Rad)
- *3: Smart Cycler is a registered trademark of Cepheid Corporation.



IV. Materials Required but not Provided

1. Reagents

- PCR primers For real-time PCR primer design, please refer to Section IX.-1.
- Sterile distilled water

2. Materials

- Real-time PCR reaction tubes or plates
- Micropipettes and tips (autoclave treated)
- Gene amplification system for real-time PCR (authorized instruments) Compatible instruments include:
 - Applied Biosystems 7300/7500/7500 Fast Real-Time PCR System, StepOnePlus Real-Time PCR System (Life Technologies)
 - LightCycler/LightCycler 480 System (Roche Diagnostics)
 - · Smart Cycler II System (Cepheid)
 - CFX96 Real-Time PCR Detection System (Bio-Rad)
 - Thermal Cycler Dice Real Time System Single (Cat. #TP850/TP870)*
 Thermal Cycler Dice Real Time System Lite (Cat. #TP700/TP760)*

Note: It is recommended that SYBR *Premix Ex Taq* II (Tli RNaseH Plus) (Cat. #RR820A) is used with the Thermal Cycler Dice Real Time System // (Cat. #TP900/TP960)*.

*: Not available in all geographic locations. Check for availability in your region.

V. Storage

- Store at 4°C (stable for up to 6 months).
- Protect this kit from light and avoid contamination.
- This kit is shipped in a dry ice-containing package. Store the kit protected from light at 4°C after receipt.
- For long-term storage, store at -80°C· (Do not store at -20°C.) Store thawed product at 4°C and use within 6 months.

VI. Precautions

Read these precautions before use and follow them when using this product.

- 1. Before use, make sure the reagent is evenly mixed by gently inverting several times without creating bubbles. Uneven reagent mixing will result in inadequate reactivity.
 - Do not mix by vortexing.
 - When stored at -80°C, SYBR *Premix Ex Taq* (2X conc.) may develop a white to pale yellow precipitant. Gently hand-warm and let stand protected from light at room temperature. Invert gently several times to dissolve the precipitate completely.
 - The presence of precipitant is indicative of uneven reagent distribution; make sure that the reagent is evenly mixed before use.
- 2. Place reagents on ice when preparing the reaction mixture.

- 3. This product contains SYBR Green I. Avoid exposure to strong light when preparing the reaction mixture.
- 4. Use fresh disposable tips to avoid contamination between samples when preparing or dispensing reaction mixtures.

VII. Protocol

Note: Please follow the procedures outlined in the manual of each respective instrument.

< Applied Biosystems 7300/7500/7500 Fast Real-Time PCR System and StepOnePlus Real-Time PCR System >

A. Prepare the PCR mixture shown below.

<Per reaction>

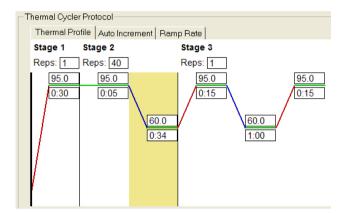
Reagent	Volume	Volume	Final conc.
SYBR <i>Premix Ex Taq</i> (Tli RNaseH Plus) (2 X)	10 μΙ	25 μΙ	1X
PCR Forward Primer (10 μ M)	0.4 μΙ	1 μΙ	$0.2 \mu M^{*1}$
PCR Reverse Primer (10 μ M)	0.4 μΙ	$1 \mu I$	$0.2 \mu M^{*1}$
ROX Reference Dye (50 X) or			
Dye II (50 X)*2	0.4 μΙ	$1~\mu$ l	1X
Template *3	2 μΙ	4 μΙ	
dH ₂ O (sterile distilled water)	$6.8 \mu I$	18 μΙ	
Total	20 μΙ*4	50 µI *4	

- *1: A final primer concentration of 0.2 μ M likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μ M.
- *2: The concentration of ROX Reference Dye II (50X) is lower than that of ROX Reference Dye (50X).
 - Use ROX Reference Dye II (50X) when performing analyses with Applied Biosystems 7500/7500 Fast Real-Time PCR System.
 - Use ROX Reference Dye (50X) when using StepOnePlus or Applied Biosystems 7300 Real-Time PCR System.
- * 3: The quantity varies depending on the number of target copies present in the template solution. Make serial dilutions to determine the appropriate template amount and use no more than 100 ng of DNA template per 20 μ l. Furthermore, if cDNA (RT reaction mixture) is used as template, the volume of the RT reaction mixture should be no more than 10% of PCR mixture.
- *4: Prepare in accordance with the recommended volume for each instrument.

B. Start the reaction.

The recommended shuttle PCR (i.e., 2-step PCR) protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform 3-step PCR when using primers with low Tm values or when a shuttle PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)

1) Applied Biosystems 7300/7500 Real-Time PCR System, StepOnePlus



Stage 1: Initial denaturation Reps: 1 95° C 30 sec. Stage 2: PCR Reps:40 95° C 5 sec. 60° C 30 - 34 sec. * Dissociation stage

*: StepOnePlus, set to 30 sec.; 7300, set to 31 sec.; 7500, set to 34 sec.

Figure 2. Shuttle PCR standard protocol.

2) Applied Biosystems 7500 Fast Real-Time PCR System

```
Shuttle PCR standard protocol
Hold Stage
Reps: 1
95°C 30 sec.
Cycling Stage
Number of Cycles: 40
95°C 3 sec.
60°C 30 sec.
Melt Curve Stage
```

Note:

- TaKaRa Ex Taq HS is a hot start PCR enzyme that includes an anti-Taq antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95°C for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95°C for 30 sec. is sufficient.
- After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.
 Refer to the instrument's instruction manual for specific analysis methods.



< LightCycler/LightCycler 480 System >

A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Final conc.
SYBR <i>Premix Ex Taq</i> (Tli RNaseH Plus) (2X)	10 μΙ	1X
PCR Forward Primer (10 μ M)	0.4 μΙ	$0.2~\mu{ m M}^{*1}$
PCR Reverse Primer (10 μ M)	0.4 μΙ	$0.2 \mu M * 1$
Template (< 100 ng)*2	2 μΙ	
dH ₂ O (sterile distilled water)	7.2 µl	
Total	20 μΙ	

- *1: A final primer concentration of 0.2 μ M likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μ M.
- *2: The optimal amount depends on the number of target copies in the template solution. Make serial dilutions to determine the appropriate amount and to use no more than 100 ng of DNA template. Furthermore, if cDNA (RT reaction mixture) is added as template, the template volume should be no more than 10% of the PCR mixture.

B. Start the reaction.

The recommended shuttle PCR (i.e., 2-step PCR) protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform 3-step PCR when using primers with low Tm values or when a shuttle PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)

<LightCycler>

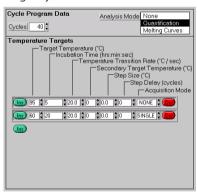


Figure 3. Shuttle PCR standard protocol.

Stage 1: Initial denaturation 95°C 30 sec. 20°C/sec. 1 cycle

Stage 2: PCR (See figure on the left) 95°C 5 sec. 20°C/sec. 60°C 20 sec. 20°C/sec. 40 cycles

Stage 3: Melt Curve Analysis 95°C 0 sec. 20°C/sec. 65°C 15 sec. 20°C/sec. 95°C 0 sec. 0.1°C/sec.

<LightCycler 480 System>

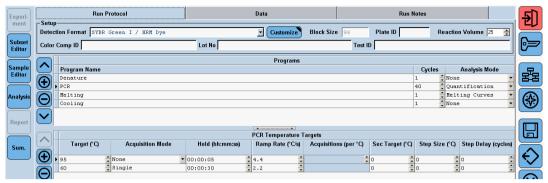


Figure 4. Shuttle PCR standard protocol

```
Denature
   95°C 30 sec. (Ramp rate: 4.4°C/sec.)
   1 cycle
PCR
   Analysis Mode: Quantification
   95°C 5 sec. (Ramp rate: 4.4°C/sec.)
   60°C 30 sec. (Ramp rate: 2.2°C/sec., Acquisition Mode : Single)
   40 cycles
Melting
   Analysis Mode: Melting Curves
   95°C 5 sec. (Ramp rate: 4.4°C/sec.)
   60^{\circ}C 1 min. (Ramp rate: 2.2^{\circ}C/sec.)
   95℃
                  (Ramp rate: 0.11°C/sec., Acquisition Mode: Continuous, Acquisitions
                   :5 per °C)
   1 cycle
Coolina
   50°C 30 sec. (Ramp rate: 2.2°C/sec.)
   1 cycle
```

Note:

- TaKaRa Ex Tag HS is a hot start PCR enzyme includes an anti-Tag antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95°C for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95°C for 30 sec. is sufficient.
- C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed. Refer to the instrument manual for specific analysis methods.



< Smart Cycler II System >

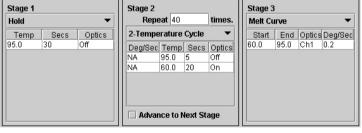
A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Final conc.
SYBR <i>Premix Ex Taq</i> (Tli RNaseH Plus) (2X)	12.5 μΙ	1X
PCR Forward Primer (10 μ M)	0.5 μΙ	$0.2 \mu M * 1$
PCR Reverse Primer (10 μ M)	0.5 μΙ	$0.2 \mu M *1$
Template ($< 100 \text{ ng}$)*2	2.0 μΙ	
dH ₂ O (sterile distilled water)	9.5 µl	
Total	25 μΙ	

- *1: A final primer concentration of 0.2 μ M likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μ M.
- *2: The quantity varies depending on the number of target copies present in the template solution. Make serial dilutions to determine the appropriate amount and use no more than 100 ng of DNA template. If cDNA (RT reaction mixture) is used as template, the volume of the RT reaction mixture should be no more than 10% of the PCR mixture.
- B. Briefly centrifuge reaction tubes with the Smart Cycler centrifuge and then set them in the Smart Cycler instrument to initiate the reaction.

The recommended shuttle PCR (i.e., 2-step PCR) protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform 3-step PCR when using primers with low Tm values or when a shuttle PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)



Stage 1: Initial denaturation Hold 95°C 30 sec. Stage 2: PCR Repeat: 40 times 95°C 5 sec. 60°C 20 sec. Stage 3: Melt curve

Figure 5. Shuttle PCR standard protocol.

Note:

- TaKaRa Ex Taq HS is a hot start PCR enzyme that includes an anti-Taq antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95°C for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95°C for 30 sec. is sufficient.
- C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.

For the analysis methods when using the Smart Cycler System, please refer to the instruction manual for Smart Cycler System.



< CFX96 Real-Time PCR Detection System >

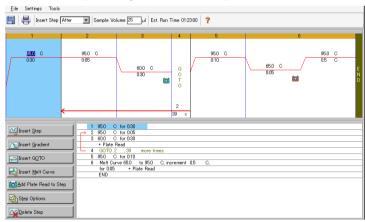
A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Final conc.
SYBR <i>Premix Ex Taq</i> (Tli RNaseH Plus) (2X)	12.5 μl	1 X
PCR Forward Primer (10 μ M)	0.5 μl	$0.2 \mu M^{*1}$
PCR Reverse Primer (10 μ M)	0.5 μΙ	$0.2 \mu M^{*1}$
Template (<100 ng)*2	2 µ1	
dH ₂ O (sterile distilled water)	9.5 µl	
Total	25 μΙ	

- *1: A final primer concentration of 0.2 μ M likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and 1.0 μ M.
- *2: The optimal amount depends on the number of target copies in the template solution. Make serial dilutions to determine the appropriate amount and use no more than 100 ng of DNA template. If cDNA (RT reaction mixture) is added as template, the template volume should be no more than 10% of the PCR mixture.
- B. Start the reaction.

The recommended shuttle PCR (i.e., 2-step PCR) protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform 3-step PCR when using primers with low Tm values or when a shuttle PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)



Sample volume: 25 μ l

Step1:

95°C 30 sec.

Step2: PCR

GOTO: 39 (40 cycles)

95°C 5 sec. 60°C 30 sec.

Step 3: Melt Curve

Figure 6. Shuttle PCR standard protocol.

Note

- TaKaRa Ex Taq HS is a hot start PCR enzyme that includes an anti-Taq antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95°C for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95°C for 30 sec. is sufficient.
- C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.

For analysis methods, refer to the manual for CFX96 Real-Time PCR Detection System.



< Thermal Cycler Dice Real Time System Single and Lite >

A. Prepare the PCR mixture shown below.

<Per reaction>

Reagent	Volume	Final conc.
SYBR <i>Premix Ex Taq</i> (Tli RNaseH Plus) (2X)	12.5 μl	1X
PCR Forward Primer (10 μ M)	0.5 μΙ	0.2 μ M *1
PCR Reverse Primer (10 μ M)	0.5 μΙ	0.2 μ M *1
Template (<100 ng)*2	2.0 μΙ	
dH ₂ O (sterile distilled water)	9.5 µl	
Total	25 μl * ³	

- *1: A final primer concentration of 0.2 μ M likely will provide good results. However, if there is an issue with reactivity, use a primer concentration between 0.1 and $1.0 \mu M.$
- *2: The optimal amount varies depending on the number of target copies present in the template solution. Make serial dilutions to determine the appropriate amount and use no more than 100 ng of DNA template. If cDNA (RT reaction mixture) is used as template, the volume of the RT reaction mixture should be no more than 10% of PCR mixture.
- *3: The recommended volume is 25 μ l for reaction mixtures.

B. Start the reaction.

The recommended shuttle PCR (i.e., 2-step PCR) protocol is described below. Use this protocol first and optimize PCR conditions when necessary. Perform 3-step PCR when using primers with low Tm values or when a shuttle PCR is not feasible. (For optimizing PCR conditions, please refer to Section VIII.)

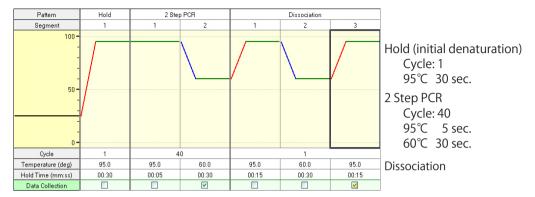


Figure 7. Shuttle PCR standard protocol.

Note:

- TaKaRa Ex Taq HS is a hot start PCR enzyme that includes an anti-Taq antibody that inhibits polymerase activity. The initial denaturation step prior to PCR should be 95°C for 30 sec. Longer heat treatment may decrease enzyme activity and affect amplification efficiency and quantification.
- For initial template denaturation before PCR, 95°C for 30 sec. is sufficient.
- C. After the reaction is complete, check the amplification and melting curves and plot a standard curve if absolute quantification will be performed.
 When using Thermal Cycler Dice Real Time System, please refer to the instruction manual for analysis methods.



VIII. Optimization

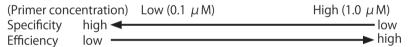
If unsatisfactory results are obtained using the recommended conditions (shuttle PCR standard protocol), follow the procedures below to optimize the primer concentration and PCR conditions. In addition, depending on the reaction system, switching to another SYBR Premix (Cat. #RR820S/A/B, RR091A/B*) may greatly improve the results.

Select PCR conditions based on comprehensive analysis of both reaction specificity and amplification efficiency. Using conditions that balance these two aspects will allow accurate measurement over a wide range of concentrations.

- *: Not available in all geographic locations. Check for availability in your region.
 - O System with a high reaction specificity
 - With no template control, non-specific amplification (e.g., primer-dimers) does not occur.
 - Non-specific amplification products, those other than the target product, are not generated.
 - System with a high amplification efficiency
 - Amplification product is detected early (small Ct value).
 - PCR amplification efficiency is high (near the theoretical value of 100%).

1. Evaluation of primer concentration

The relationship between primer concentration and reaction specificity and amplification efficiency is illustrated below. Reducing primer concentration raises reaction specificity specificities. In contrast, increasing primer concentration raises amplification efficiency.

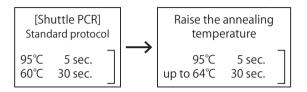




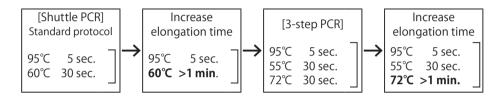


2. Evaluation of PCR conditions

○ To improve reaction specificity
Raising the annealing temperature may improve reaction specificity. Perform optimization while checking for effects on amplification efficiency.



○ To improve amplification efficiency Increasing elongation time or switching to a 3-step PCR protocol may improve amplification efficiency. Perform optimization using the steps below.



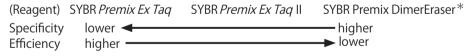
Initial denaturation

Generally, 95° C for 30 sec. is sufficient for initial denaturation, even for difficult to denature templates such as circular plasmids and genomic DNA. This step may be extended to 1 - 2 min. at 95° C depending on the template. However, prolonging this step may inactivate the enzyme. Therefore, it is recommended to avoid initial denaturation steps longer than 2 min.

3. Relationship between reagent and reactivity

Takara Bio supplies three different reagents for SYBR Green real-time PCR analysis. The relationship between reaction specificity and amplification efficiency for these reagents is described below.

SYBR *Premix Ex Taq* (Tli RNaseH Plus) (Cat. #RR420A/B) provides high amplification efficiency. SYBR *Premix Ex Taq* II (Tli RNaseH Plus) (Cat. #RR820A/B) and SYBR Premix DimerEraser™ (Perfect Real Time) (Cat. #RR091A/B)* have greater specificity.



*: Not available in all geographic locations. Check for availability in your region



IX. Appendix

1. Primer design

Designing primers with good reactivity is critical to efficient real-time PCR. Please follow the guidelines below to design primers that yield high amplification efficiency without non-specific amplification.

RT-PCR primers designed and synthesized using these guidelines are compatible with the standard shuttle PCR protocol (Section VII.).

■ Amplification product

Amplification size	The optimal size is 80 - 150 bp (amplification up to 300 bp is pos-
	sible)

Primer

Length	17 - 25 mer
GC content	40 - 60% (preferably 45 - 55%)
Tm	Make sure that the Tm values for the forward primer and the reverse primer do not differ greatly. Use primer design software to determine Tm values. OLIGO* 1 : 63 - 68°C Primer3* 2 : 60 - 65°C
Sequence	Make sure that overall there are no base sequence biases. Avoid having any GC-rich or AT-rich regions in the sequence (particularly at the 3' end). Avoid having consecutive T/C pairings (polypyrimidine). Avoid having consecutive A/G pairings (polypurine).
3' end sequence	Avoid having any GC-rich or AT-rich sequence at the 3' end. It is preferable to have a G or C as the 3' end-base. Avoid primers with T as the 3' end-base.
Complementation	Avoid having any complementary sequences of 3 bases or more within a primer and between primers. Avoid having any complementary sequences of 2 bases or more at the primer's 3' ends.
Specificity	Verify primer specificity by BLAST search *3.

- *1: OLIGO Primer Analysis Software (Molecular Biology Insights)
- *2: Primer3 (http://biotools.umassmed.edu/bioapps/primer3_www.cgi)
- *3: http://www.ncbi.nlm.nih.gov/BLAST/

2. When performing real-time RT-PCR

To synthesize cDNA templates for real-time RT-PCR, we recommend using PrimeScript™ reverse transcriptase products.

- PrimeScript RT Reagent Kit (Perfect Real Time) (Cat. #RR037A/B)
- PrimeScript RT Master Mix (Perfect Real Time) (Cat. #RR036A/B)
- PrimeScript RT Reagent Kit with gDNA Eraser (Perfect Real Time) (Cat. #RR047A/B)

When used in combination with this kit, these products can provide highly reliable results.

A. Prepare a PCR mixture by the following procedure. (When using Thermal Cycler Dice Real Time System)

Prepare the following components in volumes slightly more than that needed for the required number of tubes and dispense $22.5 - 24 \mu I$ into tubes.

<Per reaction>

Reagent	Volume	Final conc.
SYBR <i>Premix Ex Taq</i> (2X)	12.5 µl	1X
PCR Forward Primer (10 μ M)	0.5 μ l	0.2 μΜ
PCR Reverse Primer (10 μ M)	0.5 µl	0.2 μM
dH ₂ O (sterile distilled water)	xμl	
Total	22.5 - 24 µl	

B. Add 1 - 2.5 μ I of the reverse transcription reaction mixture to each of the microtubes containing aliquots of the reaction mixture.

Note: Add no more than 2.5 $\,\mu$ I of the reverse transcription reaction solution to the PCR mixture.

[Experimental example]

Human ATP5F1 mRNA was detected by real-time RT-PCR. cDNA equivalent to 1 pg - 100 ng of total RNA was used as the template, with dH₂O as the negative control.

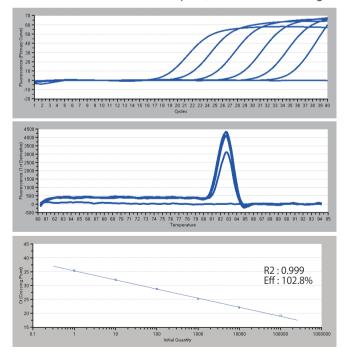


Figure 8. Detection of human ATP5F1 mRNA using real-time RT-PCR



X. Quality Specifications

1. Quality test

Consistent amplification results and SYBR Green I detection were observed for real-time PCR using λ DNA as a template (amplified fragment: 300 bp).

2. TaKaRa Ex Tag HS

A. Definition of activity

1 U is defined as the amount required to incorporate 10 nmol of total nucleotides into acid-insoluble precipitate over 30 min. at 74°C using activated salmon sperm DNA as template/primer.

Composition of the reaction mixture for assaying activity

25 mM TAPS buffer (pH9.3, 25°C)
50 mM KCI
2 mM MgCl₂
0.1 mM DTT
200 μ M each dATP, dGTP and dCTP
100 μ M [³H]-dTTP
0.25 mg/ml Activated salmon sperm DNA

B. Purity

- 1) No change in DNA electrophoresis pattern observed after 10 U of this enzyme and 0.6 μ g of λ -Hind III digest are allowed to react at 74°C for 1 hour.
- 2) No change in DNA electrophoresis pattern observed after 10 U of this enzyme and 0.6 μ g of supercoiled pBR322 DNA are allowed to react at 74°C for 1 hour.
- 3) No change in DNA electrophoresis pattern observed after 10 U of this enzyme and 0.6 μ g of λ DNA are allowed to react at 74°C for 1 hour.

XI. Related Products

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SYBR® Premix Ex Taq<sup>™</sup> (Tli RNaseH Plus) (Cat. #RR420S/A/B/L/W/LR/WR)
SYBR® Premix Ex Taq<sup>™</sup> II (Tli RNaseH Plus) (Cat. #RR820S/A/B/L/W/LR/WR)
SYBR® Premix DimerEraser<sup>™</sup> (Perfect Real Time) (Cat. #RR091A/B)*
SYBR® Premix Ex Taq<sup>™</sup> GC (Perfect Real Time) (Cat. #RR071A/B)*
PrimeScript<sup>™</sup> RT Reagent Kit (Perfect Real Time) (Cat. #RR037A/B)
PrimeScript<sup>™</sup> RT Master Mix (Perfect Real Time) (Cat. #RR036A/B)
PrimeScript<sup>™</sup> RT Reagent Kit with gDNA Eraser (Perfect Real Time) (Cat. #RR047A/B)
One Step SYBR® PrimeScript<sup>™</sup> RT-PCR Kit (Perfect Real Time) (Cat. #RR086A/B)*
One Step SYBR® PrimeScript<sup>™</sup> PLUS RT-PCR Kit (Perfect Real Time) (Cat. #RR096A/B)*
Thermal Cycler Dice<sup>™</sup> Real Time System Single (Cat. #TP850/TP870)*
Thermal Cycler Dice<sup>™</sup> Real Time System Lite (Cat. #TP700/TP760)*
Smart Cycler II System (Cat. #SC200N/SC210N)*
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*: Not available in all geographic locations. Check for availability in your region.

Cat. #RR420Q v201312Da



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[P5] PCR Notice

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[L11] SYBR® Green I

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[L15] Hot Start PCR

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[M40] Thermostable RNase H

This product is covered by the claims of U.S. Patent No. 7,422,888 and its foreign counterpart patent claims.

[M57] LA Technology

This product is covered by the claims 6-16 outside the U.S. corresponding to the expired U.S. Patent No. 5,436,149.

[M82] Tli RNaseH Plus

This product is the subject of the pending JP patent application.



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