

Make Your Own "Mate & Plate™" Library System

Catalog No. Amount Lot Number

630490 5 rxns Specified on product label.

Description

Mate & Plate Libraries are by far the easiest libraries to screen for protein-protein interactions using a GAL4 yeast two-hybrid system. Several Mate & Plate Libraries are available ready-made from Takara Bio USA, Inc. For libraries that are not available, this system provides the necessary components and a simple, highly efficient method to make your own Mate & Plate Library using SMART® technology and the highly efficient homologous recombination machinery of *Saccharomyces cerevisiae*.

Package Contents

Box 1:

- 10 μl SMART MMLV RT (200 units/μl; also available as Cat. No. 639522)
- 300 µl 5X First-Strand Buffer
- 165 µl DTT (100 mM)
- 10 μl CDS III Primer (12 μM)
- 10 μl CDS III/6 Primer (10 μM)
- 7 μl RNase H (2 units/μl)
- 50 μl 5' PCR Primer (10 μM)
- 50 μl 3' PCR Primer (10 μM)
- 500 μl Melting Solution
- 20 μl SV40 Large T PCR Fragment (25 ng/μl)
- 50 µl dNTP Mix (10 mM each dNTP)
- 25 μg pGADT7-Rec AD Cloning Vector (Sma I-linearized; 500 ng/μl)

Box 2:

- YeastmakerTM Yeast Transformation System 2 (Box 1 of 2):
- 2 x 1 ml Yeastmaker Carrier DNA, denatured (10 mg/ml)
- – 20 μl pGBT9 (100 ng/μl; control plasmid)

Box 3:

- Yeastmaker Yeast Transformation System 2 (Box 2 of 2):
- - 2 x 50 ml 50% PEG
- - 50 ml 1 M LiAc (10X)
- - 50 ml 10X TE Buffer
- 50 ml YPD Plus Liquid Medium

Box 4:

- 10 μl SMART III Oligonucleotide (12 μM)
- 5 μl Control Poly A⁺ RNA (Mouse Liver; 1 μg/μl)
- 0.5 ml S. cerevisiae Y187

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Box 5:

- 1 pouch YPDA Broth (0.5 L)
- 1 pouch YPDA with Agar (0.5 L)
- 1 pouch SD/-Leu with Agar (0.5 L)
- 50 ml NaCl Solution (0.9%)
- 300 μ1 Sodium Acetate (3 M; pH 4.8)
- 500 μl Deionized H₂O
- 10 each CHROMA SPINTM+TE-400 Columns

Storage Conditions

- Store Boxes 1 & 2 at -20° C.
- Store Box 4 at -70° C.
- Store Boxes 3 & 5 at room temperature.

Expiration Date

• Specified on product label.

Shipping Conditions

- Boxes 1, 2 & 4: Dry ice (-70°C)
- Boxes 3 & 5: Room temperature

Product Documents

Documents for our products are available for download at <u>takarabio.com/manuals</u> The following documents apply to this product:

- Make Your Own "Mate & Plate" Library System User Manual
- Yeastmaker Yeast Transformation System 2 User Manual
- pGADT7-Rec Vector Information

Quality Control Data

1. Plasmid Identity and Purity

a. The identity of the pGADT7-Rec Vector was verified by agarose/EtBr gel electrophoresis after digestion with the indicated enzyme. The purity of the vector was checked by determining the A_{260}/A_{280} .

Vector	Enzyme(s)	Fragment Size (kb)
pGADT7-Rec	Smal	8.0
A_{260}/A_{280} : 1.8–2.0		

b. The pGADT7-Rec AD Cloning Vector (SmaI-linearized) was also checked by transformation into StellarTM Competent Cells (Cat. No. 636763). Transformants were selected by plating on LB/Amp (100 μg/ml). The linearized vector produced ≤ 0.5% of the number of colonies produced with circular (uncut) pGADT7-Rec.

2. Yeast Strain

The nutritional requirements (i.e., auxotrophic markers) of the Y187 strain were verified by streaking samples onto several types of SD minimal media.

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3. cDNA Synthesis & Recombination-mediated Cloning

- a. Single-stranded cDNA was generated from 1 μg of Control Mouse Liver Poly A⁺ RNA using SMART MMLV Reverse Transcriptase as described in the User Manual (PT4085-1). Two samples of first-strand cDNA were prepared: One was generated with the CDS III Primer; the other, with the CDS III/6 Primer. Next, 2 μl of each cDNA sample was amplified by PCR, to prepare ds cDNA as described in the User Manual. Finally, 5 μl of the PCR product was electrophoresed on a 1.2% agarose/EtBr gel. A moderately strong smear from ≥0.1 kb to 4 kb (or more) was observed.
- b. Recombination-mediated cloning was tested with the following four transformations:

		ds cDNA			pGADT7-Rec2	
Transformation	Competent Y187 cells	CDS IIIa	CDS III/6 ^a	SV40b	Blank ^c	(Smal-linearized)
1.	100 μΙ	5 µl	_	_	_	1 µl
2.	100 μΙ	_	5 µl	_	_	1 µl
3.	100 μΙ	_	_	5 µl	_	1 µl
4.	100 µl		_	_	5 µl	1 µl

^a First-strand cDNA was synthesized as in Part 3a, above, using either the CDS III or CDS III/6 Primer. The cDNA was then PCR amplified, purified by gel filtration (using a CHROMA SPIN+TE-400 Column, Cat. No. 636076), and finally, concentrated by ethanol precipitation, all as described in PT4085-1.

Following transformation, the mixtures were spread on SD/-Leu plates and incubated at 30°C.

Transformation #4, a negative control, produced \leq 20% of the number of colonies observed for Transformations #1 or #2, and Transformation #3 produced \geq 4X the number of colonies observed for Transformation #4.

4. Yeast Mating

Mating functionality was confirmed by performing the following 2 matings:

- a. Y2HGold[pGBKT7-53] x Y187[pGADT7-T] (positive control mating)
- b. Y2HGold[pGBKT7-lam] x Y187[pGADT7-T] (negative control mating)

Expected colony growth was observed on SD/-Leu/-Trp media and SD/-Leu/-Trp/-Ade/-His media containing Aureobasidin A and X-alpha-Gal.

It is certified that this product meets the above specifications, as reviewed and approved by the Quality Department.

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b The SV40 Large T PCR Fragment provided with this kit.

^c The Blank was prepared and processed in the same way as the CDS III and CDS III/6 samples with one exception: Mouse Liver Poly A⁺ RNA was omitted from the first-strand synthesis reaction.



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United States/Canada Asia Pacific Europe Japan 1/24/2020